

Analysis of the Microbial Community in a Wastewater Sequencing Batch Reactor

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Abstract:

Sequencing Batch Reactors (SBRs) increase the biofilm surface area in sewage treatment units that decrease the nitrogen content of wastewater. Our hypothesis is that the species composition of the SBR biofilm will change during the course of operation. Early, denitrification-positive biofilm samples were collected after two weeks of SBR operation and 16S rRNA genes from uncultured organisms were amplified, cloned, and sequenced. One third of the clones corresponded to previously cultured Gamma Proteobacteria, Bacteroidetes, and Actinobacteria. The remaining sequenced clones were less than 95% identical to GenBank database sequences and therefore represented new genera. These were most similar to 16S rRNA sequences from uncultured Proteobacteria, Bacteroidetes, Actinobacteria, Verrucomicrobia, and the recently identified phylum-level division TM7.

Biofilm samples were also suspended in water, diluted and cultured on tryptic soy agar at different temperatures. Amplified 16S rRNA gene sequences from pure cultures showed that Gamma Proteobacteria dominated the collection, which included 49% in the class Enterobacteriales, 39% in the class Pseudomonadales, and 6% in the class Xanthomonadales. The remaining 6% of identified isolates were Firmicutes. Of the identified isolates, 52% were capable of nitrate reduction to nitrite, however, all were negative for denitrification.

The twelve week sample was also suspended in water, diluted, and cultured on TSA plates at different temperatures. Organisms cultured from the biofilm samples after twelve weeks of operation showed a greater diversity of organisms than after the two week operation time. Gamma Proteobacteria decreased to about 55% of the isolates identified. Actinobacteria made up about 18% of the sample while both Firmicutes and Beta Proteobacteria made up about 14% each. In this sample, there was a much higher number of organisms capable of reducing nitrate to nitrite and some, about 9%, were also capable of denitrifying nitrite.

GOAL of SBR: Decrease the nitrogen and organic carbon levels in treated wastewater effluent by increasing the biofilm surface area for the microbial community.

MECHANISM: Reactor undergoes Aerobic and Anaerobic cycles
Aerobic – chemolithotrophic oxidation of NH₃ and NO₂
Anaerobic – NO₃ and NO₂ reduction to N₂ gas (denitrification)

Project Goal: Evaluate the microbial community present in the SBR at two different time points.

Methods:



Obtained Biofilm Samples from SBR

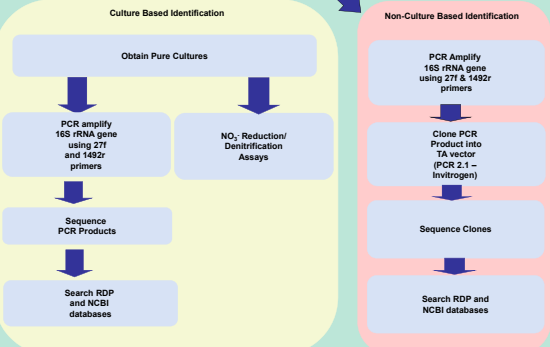


Figure 1 - Flow chart detailing the steps and procedures used in characterizing the organisms present in the biofilm at two and twelve week increments.

Results:

Table 1 – Uncultured 16S rRNA sequence identifications from biofilm after two weeks of SBR operation.

| Clone: | Sequence % Match: | Most Similar Type Strain: | Phylogeny: |
|--------|-------------------|---|--|
| BM01 | 96% | <i>Mycobacterium bonickel</i> | Actinobacteria; Mycobacteriaceae |
| BM24 | 93% | <i>Chryseobacterium aechoense</i> | Bacteroidetes; Flavobacteriaceae |
| BM07 | 88% | <i>Terrimonas ferruginea</i> | Bacteroidetes; Flavobacteriaceae |
| BM18 | 86% | <i>Terrimonas ferruginea</i> | Bacteroidetes; Flavobacteriaceae |
| BM17 | 81% | <i>Halscomenobacter hydrossis</i> | Bacteroidetes; Saprospiraceae |
| BM02 | 90% | <i>Haliangium ochraceum</i> | δ Proteobacteria; Haliangiaceae |
| BM35 | 95% | <i>Acinetobacter schindleri</i> | γ Proteobacteria; Moraxellaceae |
| BM21 | 99% | <i>Citrobacter brakii</i> | γ Proteobacteria; Enterobacteriaceae |
| BM22 | 96% | <i>Citrobacter brakii</i> | γ Proteobacteria; Enterobacteriaceae |
| BM06 | 85% | <i>Geobacter metallireducens</i> | γ Proteobacteria; Geobacteraceae |
| BM08 | 93% | <i>Xanthomonas cynarae</i> | γ Proteobacteria; Xanthomonadaceae |
| BM05 | 92% | <i>Pseudoalteromonas luteoviolacea*</i> | γ Proteobacteria; Pseudoalteromonadaceae |
| BM14 | 84% | <i>Bacillus clausii*</i> | Firmicutes; Bacillaceae |
| BM15 | 96% | <i>Clostridium baratii*</i> | Firmicutes; Clostridiaceae |
| BM19 | 88% | <i>Verrucomicrobium spinosum</i> | Verrucomicrobia; Verrucomicrobiaceae |

The * indicates that initially these bacteria were characterized in the new phylum-level division TM7. The current identifications were provided by the Ribosomal Database Project, which did not show any organisms as part of TM7. The TM7 designation actually was made using the NCBI BLAST database. This discrepancy shows the possible differences encountered when using various databases.

Table 2 – Cultured 16S rRNA sequence identifications and NO₃ reduction/ denitrification test results from biofilm after two weeks of SBR operation.

| Strain: | Sequence % Match: | Most Similar Type Strain Organism: | Phylogeny: | Test for Nitrate Reduction: | Test for Denitrification: |
|----------------------|-------------------|---|--------------------------------------|-----------------------------|---------------------------|
| BMI, K, N | 97-99% | <i>Citrobacter freundii</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BMG | 97% | <i>Enterobacter asburiae</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BMO | 98% | <i>Klebsiella granulomatis</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BM42 | 99% | <i>Klebsiella granulomatis</i> | γ-Proteobacteria; Enterobacteriaceae | - | - |
| BM32, 61 | 97-98% | <i>Klebsiella oxytoca</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BM47 | 99% | <i>Klebsiella oxytoca</i> | γ-Proteobacteria; Enterobacteriaceae | - | - |
| BM55 | 99% | <i>Klebsiella pneumoniae</i> | γ-Proteobacteria; Enterobacteriaceae | - | - |
| BMJ, M | 98-99% | <i>Klebsiella pneumoniae</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BM10 | 99% | <i>Kluyvera ascorbata</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BM12, 28 | 99% | <i>Raoultella ornithinolytica</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BM29 | 98% | <i>Raoultella planticola</i> | γ-Proteobacteria; Enterobacteriaceae | + | - |
| BMF | 97% | <i>Acinetobacter johnsonii</i> | γ-Proteobacteria; Moraxellaceae | - | - |
| BMW | 96% | <i>Acinetobacter junii novum</i> | γ-Proteobacteria; Moraxellaceae | - | - |
| BMQ, S | 98% | <i>Pseudomonas fluorescens</i> | γ-Proteobacteria; Pseudomonadaceae | + | - |
| BMR | 98% | <i>Pseudomonas fluorescens</i> | γ-Proteobacteria; Pseudomonadaceae | - | - |
| BM09, 14, 19, 23, 70 | 91-99% | <i>Pseudomonas koreensis</i> | γ-Proteobacteria; Pseudomonadaceae | - | - |
| BMP | 99% | <i>Pseudomonas rhodesiae</i> | γ-Proteobacteria; Pseudomonadaceae | + | - |
| BMA, D | 97-98% | <i>Pseudomonas umsongensis</i> | γ-Proteobacteria; Pseudomonadaceae | - | - |
| BMH | 99% | <i>Stenotrophomonas nitritireducans</i> | γ-Proteobacteria; Xanthomonadaceae | + | - |
| BMB | 99% | <i>Stenotrophomonas rhizophila</i> | γ-Proteobacteria; Xanthomonadaceae | - | - |
| BM81, E | 96% | <i>Exiguobacterium acetylucum</i> | Firmicutes; Bacillaceae | - | - |

Table 3 – Cultured 16S rRNA sequence identifications and NO₃ reduction/ denitrification test results from biofilm after twelve weeks of SBR operation.

| Strain: | Sequence % Match: | Most Similar Type Strain Organism: | Phylogeny: | Test for Nitrate Reduction: | Test for Denitrification: |
|---------------|-------------------|------------------------------------|--------------------------------------|-----------------------------|---------------------------|
| BM2.8 | 97 | <i>Microbacterium paroxydans</i> | Actinobacteria; Microbacteriaceae | + | N/A |
| BM2.9 | 95 | <i>Microbacterium novum</i> | Actinobacteria; Microbacteriaceae | - | - |
| BM2.24 | 97 | <i>Microbacterium terregens</i> | Actinobacteria; Microbacteriaceae | - | - |
| BM2.13 | 97 | <i>Rhodococcus erythropolis</i> | Actinobacteria; Nocardiaceae | - | - |
| BM2.1, 17 | 99 | <i>Bacillus pumilis</i> | Firmicutes; Bacillaceae | + | - |
| BM2.4 | 99 | <i>Bacillus licheniformis</i> | Firmicutes; Bacillaceae | + | N/A |
| BM2.15 | 98 | <i>Acidovorax defluvi</i> | β-Proteobacteria; Comamonadaceae | + | + |
| BM2.22 | 99 | <i>Acidovorax temperans</i> | β-Proteobacteria; Comamonadaceae | + | + |
| BM2.23 | 99 | <i>Comamonas testosteroni</i> | β-Proteobacteria; Comamonadaceae | - | - |
| BM 2.3 | 98 | <i>Klebsiella pneumoniae</i> | γ-Proteobacteria; Enterobacteriaceae | + | N/A |
| BM2.5 | 98 | <i>Serratia marcescans</i> | γ-Proteobacteria; Enterobacteriaceae | + | N/A |
| BM2.18 | 97 | <i>Raoultella terrigena</i> | γ-Proteobacteria; Enterobacteriaceae | + | N/A |
| BM2.19 | 98 | <i>Pantoea ananitis</i> | γ-Proteobacteria; Enterobacteriaceae | + | N/A |
| BM2.20 | 98 | <i>Pantoea agglomerans</i> | γ-Proteobacteria; Enterobacteriaceae | - | - |
| BM2.7, 11, 25 | 97-99 | <i>Aeromonas media</i> | γ-Proteobacteria; Aeromonadaceae | + | N/A |
| BM2.12 | 97 | <i>Aeromonas hydrophila</i> | γ-Proteobacteria; Aeromonadaceae | + | N/A |
| BM2.10 | 98 | <i>Pseudomonas fragi</i> | γ-Proteobacteria; Pseudomonadaceae | - | - |
| BM2.16 | 98 | <i>Pseudomonas veronii</i> | γ-Proteobacteria; Pseudomonadaceae | - | - |
| BM2.21 | 99 | <i>Pseudomonas alcaligenes</i> | γ-Proteobacteria; Pseudomonadaceae | + | N/A |

Summary:

Two Weeks: The majority of cultured organisms were γ-Proteobacteria and were representative of the 4 different families Pseudomonadaceae, Enterobacteriaceae, Xanthomonadaceae and Moraxellaceae. The only other cultured organism was identified as a Firmicute in the family Bacillaceae. γ-Proteobacteria made up about 94% of the total sample while the single Firmicute isolate made up the other 6% of the sample.

Twelve Weeks: In this sample, there was much more diversity seen in not only the family level but also at the phylum level since there are 4 phylums represented instead of two. These phylums are Actinobacteria, Firmicutes, γ-Proteobacteria and β-Proteobacteria. There are also 7 different families in the four phylums identified. These families are Microbacteriaceae, Nocardiaceae, Bacillaceae, Comamonadaceae, Enterobacteriaceae, Pseudomonadaceae and Aeromonadaceae.

Conclusions:

➤ There were a total of 4 families found after the initial two week incubation whereas there were 7 families found after the twelve week incubation period.

➤ The diversity of the SBR microbial community increased with time. This can be seen not only in the diversification of families, but also in the increased diversity in the phylum level identifications.

References:

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